MicroRNA Expression and Target Prediction in Children with Nephrotic Syndrome

Abstract

Background: Nephrotic syndrome is a common cause of kidney diseases in children. Many studies have examined the association of microRNAs playing potential roles in many pathophysiological functions. We investigated the expression pattern of the microRNAs miR-17-5P, miR-155p, miR-424-5p in children with steroid sensitive nephrotic syndrome (SSNS) and steroid resistance nephrotic syndrome (SRNS), along with the healthy subjects. **Materials and Methods:** Total RNA was isolated from the urine samples from the three groups (SSNS n = 100, SRNS n = 100, and healthy control group n = 100). Bioinformatics tools such as miRWalk and miR-Tar link were used in predicting targets for the microRNAs. Online database and g profiler software are used to evaluate the targets based on the biological functions. The expression pattern for the candidate microRNAs was carried out using quantitative real time polymerase chain reaction (RT-PCR) equipment. **Results:** miR-424 and miR-155 were upregulated in SRNS group while miR-17 was downregulated in SRNS group. miR-424-5p and miR-155p was up regulated in SRNS group while miR-17-5p was downregulated. **Conclusion:** Combined analysis of gene expression along with studied candidate microRNAs can give better understanding of the pathogenesis of childhood nephrotic syndrome.

Keywords: Expression analysis, g profile, microRNAs, miR Tar Link, miRWalk, Nephrotic syndrome

Introduction

Nephrotic syndrome is a common kidney syndrome in children.¹⁻⁴ Certain microRNAs have been experimentally validated in order to identify and select the specific function related to this condition.⁵ Almost 80% of children with idiopathic nephrotic syndrome show remission of proteinuria with corticosteroids (steroid sensitive), but nearly 40%-60% may tend to experience intermittent relapses.⁶⁻⁸ Steroid resistance denotes the absence of remission following at least 4 weeks of prednisone therapy.9 While about 30% patients with steroid resistance have an underlying genetic etiology, the exact disease mechanisms for others remain elusive. Epigenetic modification of gene expression, including expression of noncoding RNA (microRNAs), might provide an etiological clue in patients without a definitive genetic etiology. Further research is needed to fully comprehend the implications of this discovery and explore potential interventions or treatments.¹⁰ Our study focuses on idiopathic nephrotic syndrome in children with steroid sensitive nephrotic syndrome (SSNS) and steroid resistant nephrotic syndrome (SRNS).

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, transform, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

MicroRNAs (miRNAs), which regulate target genes in the post-transcriptional processes and play crucial roles in many disorders, are about 20 nucleotides in length. miRNAs have been demonstrated to function as noninvasive biomarkers in a variety of diseases, including renal disorders, in serum and urine.^{11,12}

Both microRNAs (miRNAs) and RNA binding proteins (RBPs) are crucial transcriptional regulators of gene expression. These two factors also act to reduce translation by translation repression or miRNA cleavage.¹³ They are found to be a biomarker not only in diseases such as viral infections, cardiovascular disorders, diabetes, and cancer, but also in kidney diseases.¹⁴

Kidney disease has been discovered to be significantly influenced by miR-155. It has been linked to a number of pathological processes, including fibrosis, immune response regulation, and inflammation, all of which are crucial for the onset and progression of kidney disorders.⁶ A study by Wang *et al.*, has compared expression pattern for miR-424 between the nondiabetics along with the diabetic nephropathy patients and it was observed

How to cite this article: Charmine P, Venkatesan V, Geminiganesan S, Nammalwar BR, Dandapani MC. MicroRNA Expression and Target Prediction in Children with Nephrotic Syndrome. Indian J Nephrol. 2025;35:59-63. doi: 10.25259/ jin 47 23 Pricilla Charmine¹, Vettriselvi Venkatesan², Sangeetha Geminiganesan³, Bollam Rangaswamy Nammalwar⁴, Mohanapriya Chinambedu Dandapani²

¹Faculty of Clinical Research, Departments of ²Human Genetics, ³Pediatric Medicine, Sri Ramachandra Institute of Higher Education and Research (SRIHER), Porur, Chennai, ⁴Pediatric Nephrology, Dr. Mehta's Hospital, Chetpet, Chennai, India

Corresponding author:

Mohanapriya Chinambedu Dandapani, Department of Human Genetics, SRIHER, Chennai - 600116, Tamil Nadu, India. E-mail: mohanapriya@ sriramachandra.edu.in

DOI: 10.25259/ijn_47_23



Received: 10-02-2023 Accepted: 12-10-2023 Online First: 17-06-2024 Published: 04-01-2025 that the expression of miR-424 in the kidneys of diabetic nephropathy patients was considerably higher. Additionally, inhibiting miR-424 decreased kidney damage and fibrosis in vitro, indicating that miR-424 may be involved in the onset and progression of diabetic nephropathy in people.⁷ When compared to healthy controls, the researchers discovered miR-17 was noticeably elevated in the glomeruli of patients with membranous nephropathy. miR-17 overexpression increased albuminuria and podocyte death in cultured podocytes, pointing to a potential function for miR-17 in the onset and progression of membranous nephropathy.⁸

The present study investigated the differential expression of miR-17-5p, miR-155p, miR- 424 -5p, with miR-484-5p as endogenous control in urine samples between the study groups.

Materials and Methods

The study was conducted at Sri Ramachandra Institute of Higher Education and Research and the samples were collected from the Department of Nephrology, SRIHER, Chennai, India. The children who have met the criteria were enrolled for the study are within the age group 1–12 years. The sampling for the SRNS was taken before the starting of calcineurin inhibitors and then for the SSNS group, it was taken during the relapse. The study included 300 participants, 200 cases with SSNS and SRNS, and 100 age-matched controls. Informed consent was obtained from the parents or guardians of all participants. Ethics committee approval was obtained from the Institutional Ethics committee at Sri Ramachandra Institute of Higher Education and Research [IEC-NI/21/FEB/77/2].

Study design

Patients with SSNS were enrolled during relapse prior to initiating therapy. The SRNS subjects (defined as non response to oral prednisolone at 4 weeks of therapy) were enrolled before initiating therapy with calcineurin inhibitors.

Secondary NS and familial NS were excluded.

Computational prediction for microRNAs using online database

MicroRNA prediction tools such as miRWalk and miR Target link human were used to select the specific targets for the study. The selected microRNAs were experimentally validated and analysed using the g profiler software in order to identify and select the specific function related to this disease condition.⁵

Sample collection and RNA purification

6mL of urine was collected in a sterile container. The samples are pelleted by centrifugation for 20 min at 3,000 rpm. The urinary pellet was lysed with RNA lysis buffer and it was stored in the deep freezer at 80°C for further purpose. MicroRNA isolation was carried out for all the samples from the urinary pellet by using the

MiRNeasy Mini Kit, and the procedure was performed according to the manufacturer's protocol. Urine samples were mixed thoroughly with an equal volume of miRNeasy Serum/Plasma Spike-In Control, incubated for 2-3 min at room temperature, and then analyzed and centrifuged at 12,000g for 15 min at 4°C. After transferring the upper aqueous phase to a new collecting tube, 1.5 L of 100% ethanol were added. A 2mL collection tube containing 700 L of material was pipetted into a RNeasy Min Elute spin column and spun at 8000g for 15 s at room temperature. The supernatant was discarded. The RNeasy Min Elute spin column received about 500 L of elution buffer, and it was centrifuged at the same speed. The RNeasy Min Elute spin column was filled with approximately 500 µL of 80% ethanol and centrifuged to elute the RNA.

Quantification of purified RNA

The concentrations and purity of the isolated RNA samples were quantified using NanoDrop 1000 equipment for all the samples by measuring the absorbance at 260 and 280 nm. The ratio of 1.7–2.0 and around 50 ng concentration was observed in all the purified samples denoting the good quality of RNA. All the 300 samples were checked for quality and quantity and they were found to be good and with adequate proportion for expression study.

cDNA conversion and microRNA expression using quantitative real time PCR

The cDNA conversion was performed using the Taqman advanced cDNA synthesis kit and the cycling conditions for poly A tailing is polyadenylation 37°C for 45 min, ligation 16°C for 60 min, reverse transcription 42°C for 15 min; miR-amp reaction was enzyme activation 95°C for 5 min, deanture 95°C for 3 s, annealing/extension 60°C 30 s, stop reaction 99°C for 10 min and hold at 4°C. The converted cDNA templates were further diluted to proceed for microRNA expression analysis using Rotar Gene Q equipment (Qiagen) for the selected miRNAs miR-17-5P, miR-155p, and miR-424-5P. The RT-PCR reaction components include the Taqman advanced master mix, targeted probe, and sterile water. The RT-PCR cycling condition mentioned was elaborated. Statistical (t-test) and bioinformatics analysis were performed.

Results

Target prediction analysis

The miRtarlink has revealed that miR-155p, miR-424-5p, and miR-17-5P are found to have stronger predication connected with the diseases pathogenesis and are represented in Figure 1 and miRWalk is represented in Table 1. The targeted microRNAs were also analyzed for the various pathways that are associated with the nephrotic syndrome. A detailed analysis involving various functions of candidate miRNAs has been broadly represented with their unique IDs in Table 1.



Figure 1: (a) Represents the gene *ANLN* and its targeting microRNAs. (b) Represents the microRNA (miR-424-5p) and its targeting genes.

MicroRNA expression analysis

The basic demographic characteristics are shown in Table 1. In comparison to the control group, it was observed that miR-17, miR-424, and miR-155 were elevated in SSNS and SRNS as shown in Figure 2. Further, the microRNAs were

compared within the case groups (SSNS, SRNS) to identify the fold change and found that miR-424 and miR-155 are upregualated (two-fold increase) in SRNS group while miR-17 is found to be downregulated in SRNS group as shown in Figure 3 and Table 2.

Table 1: Target Prediction for the selected microRNAs and its biological function

•		
microRNAs	Predicted Targets	
has-miR-424-5p	VEGFA	
has-miR-17-5p	IL8	
has-miR-155-5p	IL8, AGTR1, TRPS1, NR3C1	
Description	Term Id	Corrected P Value
Molecular Function		
Cytokine receptor binding	GO: 0005126	3.952×10 ⁻⁴
Signaling receptor binding	GO: 0005102	1.025×10 ⁻³
Vascular endothelial growth factor receptor binding	GO: 0005172	1.902×10 ⁻³
Biological Process		
Cell migration	GO: 0016477	1.620×10 ⁻²
Localization of cell	GO: 0051674	2.953×10 ⁻²
Cell motility	GO: 0048870	2.953×10 ⁻²
Cellular Components		
Platelet alpha granule lumen	GO: 0031093	3.905×10 ⁻²
Reactome Pathway		
Nephrin family interactions	REAC: R-HSA-37375	2.911×10 ⁻²
Wiki Pathway		
Nephrotic syndrome	WP: WP4758	9.984×10 ⁻⁴
HP		
Abnormal nephron morphology	HP: 001257	1.165×10 ⁻³
Focal segmental glomerulosclerosis	HP: 0000097	1.715×10 ⁻²
Abnormal renal glomerulus morphology	HP: 0000095	2.653×10 ⁻³
Abnormal renal glomerulus morphology	HP: 0000095	3.929×10 ⁻³
Proteinuria	HP: 0000093	5.157×10 ⁻³
Glomerular sclerosis	HP: 0000096	5.157×10 ⁻³
Abnormal urine protein level	HP: 0020129	5.649×10 ⁻³

RNA: Ribonucleic acid, VEGFA: Vascular endothelial growth factor A, AGTR: Angiotensin II Receptor Type 1, TRPS: Trichorhinophalangeal Syndrome, NR3C: Nuclear Receptor Subfamily 3 Group C Member (also known as Glucocorticoid Receptor), HP: Haptoglobin, REAC: Ras-Responsive Element-Binding Protein (also known as ATF2 - Activating Transcription Factor 2), R-HSA: Reactome Database (R) Stable identifier corresponding species (Homosapiens (HSA), WP: Wingless-related Pathway).



Figure 2: (a-f) Urinary microRNA expression of miR-155, miR-424 and miR-17 in children with SSNS and SRNS. SSNS: Steriod sensitive nephrotic syndrome, SRNS: Steriod resistant nephrotic syndrome.

Discussion

Many studies have reported the importance of microRNAs and its potential roles in apoptosis, cell differentiation and cell proliferation with the inclusion of the kidney disease. Differential expression miRNAs studies has been helpful in genome-wide profiling studies.¹⁰⁻¹⁴ In kidney injury, miR-21 and miR-155 were found to show decreased level of expression in blood and urine samples.¹⁵ By using a micro-RNA array, Luo *et al.* showed that miRNAs mir30a05p, mir-151-3p, and miR-191 showed high level of expression in urine and serum samples in NS children when compared to the healthy individuals.¹⁶

In this study, we investigated the expression of miR-17 on the pathogenesis of nephrotic syndrome. The expression

Table 2: Relative expression analysis of microRNAsbetween SSNS and SRNS

miRNA	Fold change	P value
miR-424	SSNS = 0.12	SSNS = <.0001
	SRNS = 2.80	SRNS = 0.00056
miR-17	SSNS = 2.80	SSNS = 0.001779
	SRNS = 1.72	SRNS = 0.329064
miR-155	SSNS = 1.33	SSNS = 0.179375
	SRNS = 2.95	SRNS = 0.0009

RNA: Ribonucleic acid, SSNS: Steroid sensitive nephrotic syndrome, SRNS: Steroid resistant nephrotic syndrome.

pattern observed for miR-17 was downregulated in our study, while a study by Zang *et al.* (2020), showed that mir-17-5p was found to be significantly increased in childhood nephrotic syndrome when compared with the healthy subjects.¹⁷ The miR-17 deregulation has been seen in the kidneys of nephrotic syndrome patients in numerous investigations. The authors hypothesized that elevated miR-17 expression may aid in FSGS development by encouraging podocyte damage and dysfunction.¹⁷

Our findings revealed that miR-155 expression was significantly upregulated in the glomeruli of patients with active minimal change disease (MCD), a common cause of nephrotic syndrome in children. These results are consistent with previous studies by Ramezani, *et al.*,¹⁸ and Zhang *et al.* (2020).¹⁹

Our findings are in contrast to the study by Shaffi SK, *et al.* that has investigated the expression levels of miR-424 in urine and plasma samples renal parenchymal diseases which has shown down regulation.²⁰

The limitation of the study is that the biopsy correlation with miRNA expression was not performed because our study was not focused on the kidney biopsy results; no correlation analysis was performed for the data on the kidney biopsy.

This was the first study to report the expression pattern of the microRNAs which specifically plays an important role in the podocyte junction of actin cytoskeleton. miR-424-5p and miR-155p is upregulated in SRNS group while miR-17-5p is downregulated. Combined analysis of gene



Figure 3: (a-c) Bar diagrams showing fold change in the expression levels of miR-424.,miR-17 and miR-155 among the cases. SSNS: Sensitive nephrotic syndrome, SRNS: Steroid resistant nephrotic syndrome.

expression along with the studied candidate microRNAs can give a better understanding in bringing out the knowledge in the childhood nephrotic syndrome in future studies.

Financial support and sponsorship

The study was supported by the Department of Biotechnology, Ministry of Science and Technology (Ref No: BT/PR30523/BIC/101/1121/2018).

Conflicts of interest

There are no conflicts of interest.

References

- 1. Barnett HL, Eder HA. The nephrotic syndrome. J Chronic Dis 1957;5:108-21.
- Hill AJ, Stone DE, Elliott JP, Gerkin RD, Ingersoll M, Cook CR. Management of nephrotic syndrome in the pregnant patient. J Reprod Med 2016;61:557-61.
- 3. Raina R, Krishnappa V. An update on LDL apheresis for nephrotic syndrome. Pediatr Nephrol 2019;34:1655-69.
- Dumas De La Roque C, Prezelin-Reydit M, Vermorel A, Lepreux S, Deminière C, Combe C, *et al.* Idiopathic nephrotic syndrome: Characteristics and identification of prognostic factors. J Clin Med 2018;7:265.
- Sticht C, De La Torre C, Parveen A, Gretzmi N. RWalk: An online resource for prediction of microRNA binding sites. PLoS One 2018;13:e0206239.
- 6. Carolina Tapia; Khalid Bashir Nephrotic Syndrome May 29, 2023.
- Wang G, Yan Y, Xu N, Hui Y, Yin D. Upregulation of microrna-424 relieved diabetic nephropathy by targeting Rictor through mtor Complex2/Protein Kinase B signaling. J Cell Physiol. 2019 Jul;234(7):11646-11653
- Tsuji K, Kitamura S, Wada J. MicroRNAs as biomarkers for nephrotic syndrome. Int J Mol Sci. 2020;23;22:88.

- Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, et al. MicroRNA expression profiles classify human cancers. Nature. 2005;435:834-8.
- 10. Calin GA, Croce CM. MicroRNA signatures in human cancers. Nat Rev Cancer 2006;6:857-66.
- 11. Cameron JS. Nephrotic syndrome in the elderly. Semin Nephrol 1996;16:319-29.
- Kirk N Campbell, James A Tumlin. Protecting podocytes: A key target for therapy of focal segmental glomerulosclerosis. Am J Nephrol 2018;47(Suppl 1):14-29.
- Haas M, Meehan SM, Karrison TG, Spargo BH. Changing etiologies of unexplained adult nephrotic syndrome: A comparisonof renal biopsy findings from 1976–1979 and 1995– 1997. Am J Kidney Dis 1997;30:621-31.
- 14. Tsuji K, Kitamura S, Wada J. MicroRNAs as biomarkers for nephrotic syndrome. Int J Mol Sci 2021;22:88.
- Saikumar J, Hoffmann D, Kim T-M, Gonzalez VR, Zhang Q. Expression, circulation, and excretion profile of microRNA-21, -155, and -18a following acute kidney injury. Toxicol Sci 2012;129:256-67.
- Luo Y, Wang C, Chen X, Zhong T, Cai X, Chen S, *et al.* Increased serum and urinarymicroRNAs in children with idiopathic nephrotic syndrome. Clin Chem 2013;59:658-66.
- Zhang Y.R., Wu Y.F., Wang H., Lin X.M., Zhang X.M. Role of microRNA-17-5p in the pathogenesis of pediatric nephrotic syndrome and related mechanisms. Zhongguo Dang Dai Er Ke Za Zhi. 2020;22:958–963.
- Ramezani A, Devaney JM, Cohen S, Wing MR, Scott R, Knoblach S, *et al.* Circulating and urinary microRNA profile in focal segmental glomerulosclerosis: a pilot study. Eur J Clin Invest. 2015;45:394-404.
- 19. Zhang W, Li X, Tang Y, Chen C, Jing R, Liu T. miR-155-5p Implicates in the pathogenesis of renal fibrosis via targeting SOCS1 and SOCS6. Oxid Med Cell Longev.2020:6263921.
- Shaffi SK, Galas D, Etheridge A, Argyropoulos C. Role of MicroRNAs in renal parenchymal diseases-a new dimension. Int J Mol Sci. 2018;19:1797.