# **Original Article**

# Catheter-related Infections and Microbiological Characteristics in Coiled Versus Straight Peritoneal Dialysis Catheters in Malaysia

### Abstract

Background: Catheter-related infections remain a threat in peritoneal dialysis (PD) patients. Attempts to improve catheter insertion techniques and catheter type with best infectious outcomes yield heterogenous results. We seek to determine catheter-related infections in two different types of catheters and its microbiological spectrum. Methods: Retrospective cross-sectional study conducted in Hospital Serdang, Malaysia. We included end-stage renal disease (ESRD) patients who opted for PD and examined catheter-related infections (peritonitis, exit site infection, and tunnel tract infection) and organisms causing these infections. Results: We included 126 patients in this study; 75 patients received the coiled PD catheter (59.5%) and 51 patients received the straight PD catheter (40.5%). The majority of patients were young, under the age of 65 years old (77.3% and 72.5%) in the coiled and straight PD catheter group, respectively, and the main cause of ESRD was diabetes mellitus in both groups (78.7% vs. 92.2%). The demographic and anthropometric data were similar between both groups. Peritonitis rate (0.29 episodes/patient-years vs. 0.31 episodes/patient-years, P value = 0.909), exit site infection rate (0.31 episodes/patient-year vs. 0.37 episodes/patient-year, P value = 0.730), and tunnel tract infection rate (0.02 episodes/patient-year, P value = 0.430) were similar in the coiled versus straight PD catheter groups. The predominant organism causing peritonitis was the gram-negative organism; Escherichia coli and Klebsiella pneumoniae. In exit site and tunnel tract infections, there is a predominance of gram-negative organisms; Pseudomonas aeruginosa and K. pneumoniae. Conclusions: There was no difference in infectious outcomes between the two different types of catheters. Type of organism in both groups was gram-negative.

**Keywords:** Exit site infection, microbiological spectrum, peritoneal dialysis, peritonitis, tunnel tract infection

# Introduction

Peritoneal dialysis (PD) is becoming a more popular choice of renal replacement therapy (RRT) not only owing to its renal protective effects<sup>[1,2]</sup> but also conferring patient empowerment and autonomy.<sup>[3,4]</sup> Despite improvements in insertion techniques and catheter designs, catheter-related infection continues to be a great source of morbidity and remains a barrier to a successful PD program. This includes peritonitis, exit site infection (ESI) as well as tunnel tract infections (TTI).

While a lot of attention is being spent on the natural history and causative organisms in peritonitis, ESI and TTI impose a similar threat to a PD program. Understanding the microbial spectrum of organisms that occur within these catheter-related infections is important as geographical variations are common worldwide. Therefore, developing a guideline based on local epidemiological and microbial data will empower health-care workers to successfully treat these infections.

Existing evidence suggested no difference in infectious complications between different types of catheters. <sup>[5-8]</sup> However, most of these data are limited by its retrospective nature and small sample size. Furthermore, there is a lack of data comparing the microbial spectrum between the two types of catheter. In this study, we evaluated the different catheter-related infections in PD patients as well as its microbiological characteristics in our center as a means of continuous quality improvement to strengthen our center's success in PD programs.

**How to cite this article:** Abdul Rashid AM, Lim CT. Catheter-related infections and microbiological characteristics in coiled versus straight peritoneal dialysis catheters in Malaysia. Indian J Nephrol 2021;31:511-5.

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Received: 26-05-2020 Revised: 09-07-2020 Accepted: 23-07-2020 Published: 11-11-2020

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# Method

This is a retrospective study conducted in Hospital Serdang, Malaysia, which is a tertiary center that specializes in Nephrology. We included 126 end-stage renal disease (ESRD) patients who required catheter insertion for PD between August 2008 and December 2010. They were assigned to receive a coiled PD catheter versus a straight PD catheter based on the discretion of treating nephrologist during initial assessment according to patients' suitability. The incidence of catheter-related infections which includes peritonitis, ESI, and TTI for these patients were collected in the period of 124 months, from August 1, 2008 to December 30, 2018.

All PD catheters were inserted by a nephrologist using the peritoneoscopic method under conscious sedation, where intravenous (IV) sedation and analgesia such as midazolam and fentanyl are given during the procedure. The catheters utilized were doubled cuffed, coiled, and straight PD catheters at 57.5 cm and 47 cm, respectively. Standard catheter care with mupirocin cream and povidone iodine were employed and IV cefazolin was given as prophylactic antibiotics. Ambulatory PD was delayed for at least 2 weeks after insertion.

A standardized data collection sheet to record patient details, comorbidities as well as the occurrence of infection (peritonitis, ESI, and TTI) was used, and data were retrieved from our computerized system by trained medical personnel. The primary outcome of this study was catheter-related infections in PD patients, which include peritonitis, ESI, and TTI infection rates. We also studied the causative organism causing these infections to examine whether the type of catheter causes different microbiological characteristics in our patients.

Peritonitis was defined as presence of at least two out of three criteria: (i) signs and symptoms of peritonitis, (ii) cloudy dialysate with white cell count of 100/µL, or (iii) demonstration of organism by PD fluid culture. A peritonitis that occurred within 4 weeks of the previous episode of peritonitis was considered a relapse, thus not classified as a new infection. ESI was the presence of purulent discharge, with or without ervthema of the skin at the catheter-epidermal interface.<sup>[9]</sup> TTI was the presence of clinical inflammation or ultrasonographic evidence of collection along the catheter tunnel.<sup>[9]</sup> If the diagnosis of peritonitis was fulfilled, PD fluid sample was taken and sent to the laboratory for culture and sensitivity. For ESI and TTI, a swab and blood sample were taken and sent to the laboratory for examination of microbiological spectrum and organism sensitivity.

Inclusion criteria were all patients above the age of 18 with a diagnosis of ESRD who opted for PD (continuous ambulatory PD and automated PD). Patients who had a PD catheter for intermittent PD while awaiting vascular

access or those who were referred from another center were excluded. This study was reviewed and approved by the Medical Research and Ethics Committee of Health Ministry of Malaysia (NMRR-18-865-41205) on July 12, 2018.

Data entry and analyses were done using SPSS version 20, on an intention-to-treat basis. Numerical variables were checked for normality distribution, and appropriate measures of central dispersion were used to describe the data. It was presented in mean (SD), and independent *t*-test was used to compare means of two groups. Categorical variables were presented in frequencies and percentages. Chi-square test was used to examine associations. A *p* value of <0.05 level of significance was considered.

# **Results**

#### **Patient characteristics**

We included 126 patients in this study; 75 patients received the coiled PD catheter (59.5%) and 51 patients received the straight PD catheter (40.5%). The majority of patients were young under the age of 65 years old (77.3% and 72.5%) in the coiled and straight PD catheter group, respectively. The mean age between the two groups was similar,  $49.9 \pm 16.79$  years in the coiled PD catheter group and  $53.4 \pm 14.67$  years in the straight PD catheter group (*P*-value = 0.173). In both groups, the majority of patients were male (53.3% vs. 56.9%, *P* value = 0.696). The anthropometry data were similar between both coiled and straight PD catheter group. The main cause of ESRD was diabetes mellitus in both groups (78.7% vs. 92.2%, *P* value = 0.049). Table 1 represents the patients' demographical characteristics.

#### Peritonitis and causative organisms

In the coiled PD catheter group, the peritonitis rate was 0.29 episodes/patient-years and in the straight PD catheter group the peritonitis rate was 0.31 episodes/ patient-years (P-value = 0.909). There were 48 episodes of peritonitis in the coiled PD catheter group and 41 episodes of peritonitis in the straight PD catheter group [Table 2]. In the coiled PD catheter group, gram-negative organisms were grown in 22 isolates (45.8%), most commonly E. coli, accounting for 16.7%. Eleven episodes (22.9%) grew gram-positive organisms, the most common isolate was Methicillin Sensitive Staphylococcus aureus accounting for 6.3%. There were two episodes of fungal peritonitis (4.2%), growing mostly *Candida albicans* (C. albicans). Thirteen episodes (27.1%) were culture negative peritonitis [Figure 1a]. In the straight PD catheter group, the proportion of gram-positive and gram-negative organisms causing peritonitis was similar, 16 episodes each (39%). However, the most common gram-negative organism was K. pneumoniae, accounting for seven (17.1%)while the most common gram-positive organism was methicillin-resistant Staphylococcus epidermidis, accounting

Organism

MSSA (GP)

MRSA (GP)

MRSE (GP)

negative (GP)

No growth (NG)

Staphylococcus coagulase

Streptococcus pneumoniae (GP)

Streptococcus pyogenes (GP)

*Streptococcus viridans* (GP)

Table 1: Patient demographical characteristics			
	Coiled ( <i>n</i> =75)	Straight (n=51)	Р
Age (years)	49.4 (16.79)	53.4 (14.67)	0.173
Male	40	29	0.696
Race			
Malay	42	31	0.034
Chinese	21	19	
Indian	12	1	
Height (cm)	157.2 (10.88)	158.6 (9.5)	0.438
Weight (kg)	58.0 (14.10)	60.4 (12.49)	0.349
Body mass index (kg/m <sup>2</sup> )	23.18 (5.32)	23.1 (4.02)	0.946
Cause of ESRD <sup>a</sup>			
Diabetes mellitus	59	47	0.049
Hypertension	3	2	1.000
Glomerulonephritis	10	0	0.006
Obstructive uropathy	1	1	1.000
Unknown	1	1	1.000

<sup>a</sup>End-stage renal disease

for four episodes (9.8%). The fungal peritonitis was similar to the coiled catheter group, two episodes (4.9%) growin C. albicans and culture-negative peritonitis constitute seven episodes (17.1%) [Figure 1b].

#### Exit site infections and Tunnel tract infections

The ESI rate was similar between both group 0.31 episodes/patient-year in the coiled PD cathete group and 0.37 episodes/patient-year in the straight P catheter group (P-value = 0.730). However, the number of surgical interventions (deroofing of ESI) require in the coiled PD catheter group was higher than in th straight PD catheter group, 16 (31.4%) versus 8 (10.7%) respectively (P-value = 0.005). In both groups, we observe higher episodes of gram-negative organisms, 21 (51.2% versus 20 (52.6%) in the coiled and straight PD cathete groups, respectively [Figure 2a and 2b]. The most commo gram-negative organism was P. aeruginosa followed b K. pneumoniae. There was no fungal organism isolated the straight PD catheter group, while the coiled PD catheter group has one episode (2.4%) of fungal organism, which grew C. albicans. There were 10 episodes (24.4%) versus 8 episodes (21.1%) of culture negative ESI in the coiled and straight PD catheter group, respectively [Table 3].

The rate of TTI was similar between both groups, 0.02 episodes/patient-year (*P*-value = 0.430). The majority of TTIs were caused by gram-negative organisms, 2 (50%) in each group. Table 4 shows the causative organisms causing TTI in both groups.

#### **Discussion**

The most common catheter-related complication is attributed to infections, where peritonitis accounts for 61%, while ESI and TTI make up 23%.<sup>[10]</sup> This is not surprising Fungal

No growth

				0				
gure	1: (a)	Organisms	causing	peritonitis	in	the	coiled	PD
theter	r and (b	) organisms c	ausing per	itonitis in the	stra	ight	PD cath	eter

Gram

Fungal

5%

No growth 17%

Positive

Streptococcus agalactiae (GP) Streptococcus salivarius (GP) Streptococcus parasanguinis (GP) Group B Streptococcus (GP) Bacillus cereus (GP) Klebsiella pneumoniae (GN) Klebsiella ozaenae (GN) ESBL Klebsiella pneumoniae (GN) Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN)	$ \begin{array}{c} 1\\ 0\\ 0\\ 1\\ 1\\ 1\\ 1\\ 8\\ 1\\ 1\\ 1\\ 2\\ 1\\ 1 \end{array} $	0 1 1 0 7 0 1 4 0 3 0 1
Streptococcus salivarius (GP) Streptococcus parasanguinis (GP) Group B Streptococcus (GP) Bacillus cereus (GP) Klebsiella pneumoniae (GN) Klebsiella ozaenae (GN) ESBL Klebsiella pneumoniae (GN) Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	$\begin{array}{c} 0 \\ 0 \\ 1 \\ 1 \\ 1 \\ 1 \\ 1 \\ 8 \\ 1 \\ 1 \\ 1 \\ 2 \\ 1 \end{array}$	1 1 0 7 0 1 4 0 3 0 1
Streptococcus parasanguinis (GP) Group B Streptococcus (GP) Bacillus cereus (GP) Klebsiella pneumoniae (GN) ESBL Klebsiella pneumoniae (GN) Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	0 1 1 1 1 1 8 1 1 1 2 1	1 0 7 0 1 4 0 3 0 1
Group B Streptococcus (GP) Bacillus cereus (GP) Klebsiella pneumoniae (GN) Klebsiella ozaenae (GN) ESBL Klebsiella pneumoniae (GN) Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	1 1 1 1 8 1 1 1 2 1	1 0 7 0 1 4 0 3 0 1
Bacillus cereus (GP) Klebsiella pneumoniae (GN) Klebsiella ozaenae (GN) ESBL Klebsiella pneumoniae (GN) Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	1 1 1 8 1 1 1 2 1	0 7 0 1 4 0 3 0 1
Klebsiella pneumoniae (GN) Klebsiella ozaenae (GN) ESBL Klebsiella pneumoniae (GN) Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	1 1 8 1 1 1 2 1	7 0 1 4 0 3 0 1
Klebsiella ozaenae (GN) ESBL Klebsiella pneumoniae (GN) Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	1 1 8 1 1 1 2 1	0 1 4 0 3 0 1
ESBL Klebsiella pneumoniae (GN) Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	1 8 1 1 1 2 1	1 4 0 3 0 1
Escherichia coli (GN) Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	8 1 1 2 1	4 0 3 0 1
Enterobacter cloacae (GN) Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	1 1 2 1	0 3 0 1
Pseudomonas aeruginosa (GN) Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	1 1 2 1	3 0 1
Burkholderia pseudomallei (GN) Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	1 2 1	0
Serratia marcescens (GN) Proteus mirabilis (GN) Citrobacter freundii (GN)	2 1	1
Proteus mirabilis (GN) Citrobacter freundii (GN)	1	
Citrobacter freundii (GN)		0
	2	0
Citrobacter kaferii (GN)	1	0
Acinetobacter baumannii (GN)	1	0
Shewanella putrefaciens (GN)	1	0
Candida albicans (fungal)	2	2
Total NG	13 (27.1)	7 (17.1)
Total GP	11 (22.9)	16 (39)
Total GN	22 (45.8)	16 (39)
Total fungal	22(13.0)	2(4.9)
Total episodes	48 (100)	41 (100)

Table 2: The causative organisms causing peritonitis in both groups

Coiled

n (%)

13 (27)

3

1

2

0

0

0

1

Straight

n (%)

7 (17.1)

2

2

4

1

1

1 2

0

grou	ips	,
Organism	Coiled <i>n</i> (%)	Straight n (%)
No growth (NG)	10 (24.4)	8 (21.1)
MSSA (GP)	5	4
MRSA (GP)	1	0
MRSE (GP)	3	3
Enterococcus faecalis (GP)	0	1
Micrococcus luteus (GP)	0	1
GPC (GP)	0	1
Klebsiella pneumoniae (GN)	5	2
Escherichia coli (GN)	0	3
ESBL Escherichia coli (GN)	1	0
Enterobacter cloacae (GN)	1	1
Pseudomonas aeruginosa (GN)	7	7
Pseudomonas putida (GN)	0	1
Burkholderia cepacia (GN)	0	1
Serratia marcescens (GN)	2	1
Proteus mirabilis (GN)	1	0
Citrobacter freundii (GN)	0	1
Acinetobacter baumannii (GN)	1	1
Providencia rettgeri (GN)	0	1
Sphingomonas paucimobilis (GN)	1	0
Morganella morganii (GN)	1	0
GNR (GN)	1	1
Candida albicans (fungal)	1	0
Total NG	10 (24.4)	8 (21.1)
Total GP	9 (22)	10 (26.3)
Total GN	21 (51.2)	20 (52.6)
Total fungal	1 (2.4)	0 (0)
Total episodes	41 (100)	38 (100)

Table 3:	The causative	organisms	causing	ESI in	both
		grouns			

NG-no growth, GN-gram-negative, GP-gram-positive,

MSSA-methicillin-sensitive Staphylococcus aureus,

MRSA-methicillin-resistant Staphylococcus aureus,

MRSE-methicillin-resistant Staphylococcus epidermidis,

GNR-gram-negative rod, GPC-gram-positive cocci

Table 4: The causative organisms causing TTI in bothgroups			
No growth (NG)	1	1	
MSSA (GP)	1	1	
Klebsiella pneumoniae (GN)	1	1	
Pseudomonas aeruginosa (GN)	0	1	
ESBL Escherichia coli (GN)	1	0	
	Total NG: 1	NG: 1	
	Total GP: 1	GP: 1	
	Total GN: 2	GN: 2	
	Total Fungal: 0	Fungal: 0	
Total episodes	4	4	

NG-no growth, GN-gram-negative, GP-gram-positive

as ESRD patients are chronically immunosuppressed due to chronic inflammation and uremia,<sup>[11]</sup> rendering them susceptible to infections. While a lot of studies attempted to determine the best insertion technique and catheter



Figure 2: (a) Organisms causing exit site infection in the coiled PD catheter and (b) organisms causing exit site infection in the straight PD catheter

type to improve infectious outcomes, the results are heterogenous.<sup>[5-8,12]</sup> Thus, current guidelines do not favor either method of insertion or specific catheter designs when determining the best PD access.<sup>[13,14]</sup> Instead, an emphasis has been made to encourage each center to have a dedicated team for PD, developing clear protocols for perioperative measure as well as adequate patient training and education as essentials in reducing overall incidence of peritonitis.<sup>[15]</sup>

In our study, the peritonitis rate in coiled and straight PD catheter is similar (0.29 episodes/patient-year vs. 0.31 episodes/patient-year). The most commonly isolated organism seen in both groups was from the gram-negative strain, *E. coli* and *K. pneumoniae*. Overall, gram-negative infections were more common in this series. It is contrary to worldwide reports,<sup>[16-20]</sup> except a few. As such, Prasad *et al.*<sup>[21]</sup> described a predominance of gram-negative organisms as the cause of peritonitis in India, reporting a worse outcome. Another retrospective study in Sarawak, Malaysia, reported similar occurrence of gram-negative and gram-positive organisms causing peritonitis, with trend of increasing gram-negative infection with time, causing higher rate of catheter loss.<sup>[22]</sup>

Both of these organisms are commensals of the normal bowel flora that contaminated the sterile peritoneal cavity. We found that most of our patients had a breach in sterile procedure and were not practicing good hand hygiene; thus, these organisms are most possibly of the fecal origin. Hygiene is still a major problem in developing countries where most of our patients were from the working class and lower middle class. This also explained our findings of gram-negative organisms originating from the soil, including *Serratia marcescens*, *Proteus mirabilis*, and *Citrobacter freundii*.

In our study, the coiled catheter group had more gram-negative peritonitis as compared to straight group. Another possible mechanism of infection of these gram-negative bowel floras is from the transmural migration. Several studies have noted a significant trend of tip migration and catheter dysfunction in the coiled PD catheter <sup>[7,23]</sup> that could have aggravated inflammatory

process, thus promoting transmural bacteria migration. The straight catheter group on the other hand had more gram-positive peritonitis, mostly from the *Staphylococci* species that is likely due to touch contamination of organisms from the cutaneous origin.

ESI rate was similar in both groups (0.31 episodes/ patient-year vs. 0.37 episodes/patient-year, P value = 0.730). Interestingly, the most common organism causing ESI in both groups was P. *aeruginosa*, a commonly found pathogen in hospital-acquired infections. This could be explained by the fact that ESIs in our hospital were treated with deroofing, which involved surgical procedure to expose and shave the external cuff.<sup>[9]</sup> As this procedure required use of the operation theater, most of our patients required admission to the ward, thus increasing the risk to hospital-acquired infections.

TTI rate was similar in both groups (0.02 episodes/ patient-year in both groups, P value = 0.430). Gram-negative organisms causing peritonitis and ESI were also found to cause more TTI, namely, *K. pneumoniae* and *P. aeruginosa*.

# Conclusion

There was no difference in infectious outcomes between the two different types of catheters. The spectrum of organism is also similar in both groups of catheters, which are of the gram-negative group.

#### Financial support and sponsorship

Nil.

### **Conflicts of interest**

There are no conflicts of interest.

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